

## Chemical Exchange Saturation Transfer MRI: Sensitive to Intracellular pH Change Over Time in a Rat Model of Brain Cancer

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- In biological systems, robust maintenance of pH is a fundamental homeostatic process.
- The regulation of pH gradient across the cell membrane is crucial for many cellular functions and that is altered in cancer cells.



### **OBJECTIVE**

To examine how pH<sub>i</sub> changes over time in the brain of a rat C6 glioma model using CEST-MRI

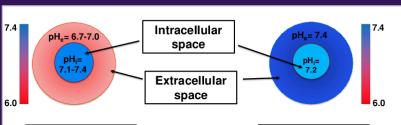


Fig.1: Schematic of intracellular pH (pH<sub>i</sub>) and extracellular pH (pH<sub>e</sub>) in normal and cancers cells

Cancer Cell Normal Cell

## **CHEMICAL EXCHANGE SATURATION TRANSFER (CEST)**

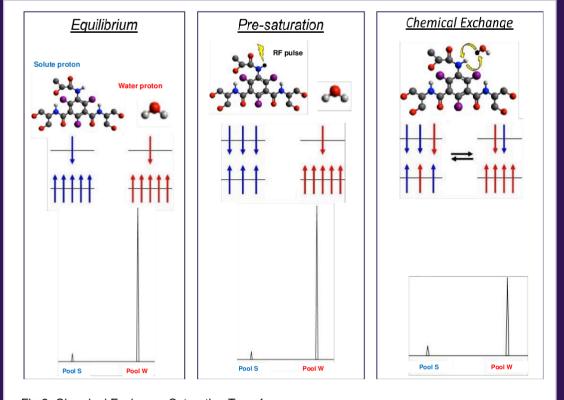
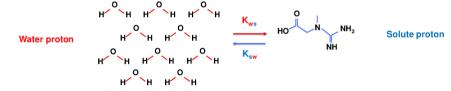


Fig.2: Chemical Exchange Saturation Transfer

Sources of funding and affiliations:

## In CEST-MRI, contrast is produced by selectively exciting exchangeable protons and detecting this exchange through the transfer of magnetization to bulk water.

- To observe the effect of saturation on a pool of dilute protons, the protons in these systems must be in exchange with one another and have a distinct chemical shift difference.
- These exchangeable protons are commonly found in amide and amine groups on mobile proteins within the intracellular space.



 The magnitude of CEST contrast depends on the proton exchange rate which is pH-dependent for amine and amide protons.

$$K_{ex} = K_0 + K_a[H^+] + K_b[OH^-] = K_0 + K_a \ 10^{-pH} + K_b \ 10^{pH-pK_w}$$

$$K_{ex} = K_b \ 10^{pH-pK_w}$$

$$K_w = [H^+][OH^-]$$
$$pK_w = -\log(K_0)$$

 $\mathbf{K_0}$ : rate constant of the spontaneous proton exchange reaction between water and the amide nitrogen nucleus.  $\mathbf{K_a}$ : rate constant of the acid-catalyzed protonation of the amide nitrogen nucleus.  $\mathbf{K_b}$ : rate constant of the base-catalyzed proton exchange reaction between hydroxyl ion and the amide nitrogen nucleus.  $\mathbf{pK_w}$ : ionization constant of water (15.4 at 37 °C).  $\mathbf{K_w}$ : dissociation constant of water.

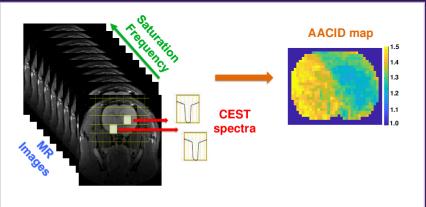
#### **EXPERIMENTAL METHODS**



Fig.3: RF pulse sequence

- B₀: 9.4 T Bruker
- Pulse sequences:
- EPI: TR= 7 s , TE= 25 ms
- FSE: TR= 7 s , TE= 7 ms FOV= 38.4×38.4 mm<sup>2</sup>
- Cells: 8µL C6 GBM
- 8-week old male Wistar rats
- Tumour position: Right frontal lobe
- Saturation frequencies: 1.2 ppm 6.6 ppm

# AACID : Amine and Amide Concentration-Independent Detection



 $AACID = \frac{M_z(3.50 \ ppm) \times [M_z(6.00 \ ppm) - M_z(2.75 \ ppm)]}{M_z(2.75 \ ppm) \times [M_z(6.00 \ ppm) - M_z(3.50 \ ppm)]}$ 

#### RESULTS

Fig.5: Average

AACID values (a) in

right and left brain of

contralateral brain of rats, measured on

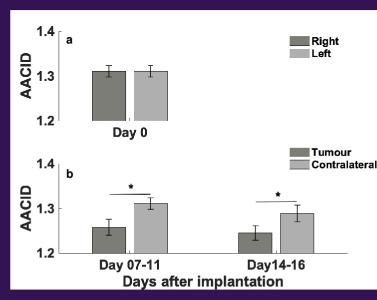
day 7-11, and day

14-16 post implantation of

Bars represent standard error of the

tumour.

control rats, (b) in tumor and



## RESULTS

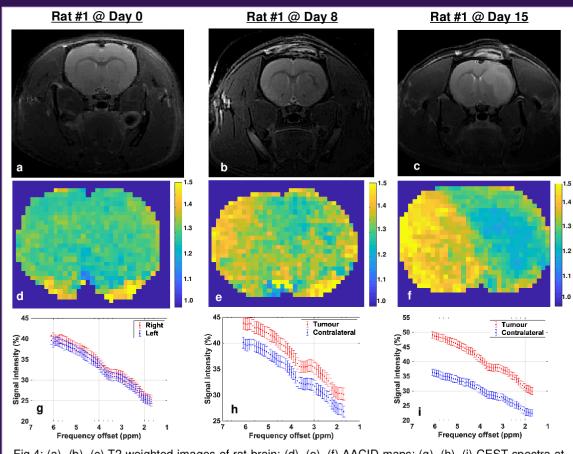


Fig.4: (a), (b), (c) T2-weighted images of rat brain; (d), (e), (f) AACID maps; (g), (h), (i) CEST spectra at day 0 and days 8 and 15 post implantation of C6 glioma cells. Purple line is the outline of the tumour region.

#### **DISCUSSION**

- No difference observed in AACID value between left and right side of the brain in control animals (N=5).
- A significantly lower average AACID value (p<0.05, paired t-test) was found in the tumour compared to the contralateral region at both time points.
- At day 7-11: average AACID value was ~4.0% lower in tumour indicating a 0.21 higher pH (N=14).
- At day 14-16: average AACID value was ~3.4% lower in tumour indicating a 0.17 higher pH (N=13).
- As expected, in this rat model, the difference between tumour pH and contralateral pH remains the same across time.

## **FUTURE STUDIES**

- Measurement of pH<sub>i</sub> by CEST following drug injection to modulate pH in C6 glioma model.
- Utilize DTI in combination with CEST to examine the effect of cellularity on pH measurements.

#### TAKE-HOME MESSAGE

- We successfully generated AACID maps of the brain in a rat C6 glioma model and demonstrate pH differences over time.
- This technique has the potential to monitor pH<sub>i</sub> in humans as no exogenous contrast agent is required.







